Mounting evidence points to LDL oxidation as an important etiologic agent of atherosclerosis. LDL is believed to exit the lumen of arteries and become trapped in the subendothelial space, perhaps by the binding of apoB-100 to intimal proteoglycans. Once trapped, LDL may become oxidized directly by cellular byproducts of respiration or enzymatically by lipoxygenases, myeloperoxidase, NADPH oxidase, or byproducts of respiratory chain. LDL oxidation is responsible in part for monocyte/macrophage recruitment and differentiation and, hence, the initiation of atherogenesis.

Substantial epidemiological evidence points to an inverse correlation between HDL levels and coronary artery disease. One plausible hypothesis explaining this phenomenon is based on the idea that HDL can exert a direct antiatherogenic effect at least in part through preventing LDL oxidation. Serum paraoxonase (PON1) is a 45-kDa glycoprotein that is expressed in the liver and has been found to be associated with HDL particles in the blood. PON1 was initially identified for its ability to hydrolyze organophosphate insecticides. More recently, PON1 has been shown to prevent LDL oxidation in vitro, and decreased levels of PON1 are associated with an increased risk of cardiovascular disease. In studies with PON1 knockout (KO) mice, PON1 has been shown to be both necessary and sufficient for the in vitro protective effects of HDL on LDL oxidation and monocyte transmigration. Furthermore, when both dietary and apoE-null models were used, PON1 KO mice exhibited an approximate 2-fold increase in atherosclerosis.

The present study aims to provide additional in vivo evidence showing that PON1 protects against atherosclerosis. In particular, we have addressed whether in an in vivo setting PON1 influences atherosclerosis in a dose-dependent manner.

**Methods**

Cloning and Microinjection of Human PON1 Gene

A clone of the human PON1 gene in a bacterial artificial chromosome vector (GenBank No. AC004022) was purchased from Genome Systems. Polymerase chain reaction (PCR) and restriction enzyme digestion confirmed that this clone contained leucine and glutamine at codons 55 and 192, respectively. This clone, designated pPON1, was purified by CsCl gradient centrifugation. The pPON1 bacterial artificial chromosome DNA was digested with Pmel and then run out on a pulse field gel to separate a 45-kb DNA fragment containing the human PON1 gene from the rest of the plasmid. The 45-kb DNA fragment was isolated from the gel, diluted to a concentration of 0.5 μg/ml, and microinjected into fertilized C57BL/6j (B6) mouse eggs (Jackson Laboratories, Bar Harbor, Me) to produce transgenic (Tg) mice.

**Background**—Serum paraoxonase (PON1), an enzyme carried on HDL, inhibits LDL oxidation, and in human population studies, low PON1 activity is associated with atherosclerosis. In addition, PON1 knockout mice are more susceptible to lipoprotein oxidation and atherosclerosis. To evaluate whether PON1 protects against atherosclerosis and lipid oxidation in a dose-dependent manner, we generated and studied human PON1 transgenic mice.

**Methods and Results**—Human PON1 transgenic mice were produced by using bacterial artificial chromosome genomic clones. The mice had 2- to 4-fold increased plasma PON1 levels, but plasma cholesterol levels were unchanged. Atherosclerotic lesions were significantly reduced in the transgenic mice when both dietary and apoE-null mouse models were used. LDL isolated from the transgenic mice also protected against LDL oxidation more effectively.

**Conclusions**—Our results indicate that PON1 protects against atherosclerosis in a dose-dependent manner and suggest that it may be a potential target for developing therapeutic agents for the treatment of cardiovascular disease. (Circulation. 2002;106:484-490.)

**Key Words:** antioxidants ■ lipoproteins ■ free radicals ■ atherosclerosis ■ genes

---

**Basic Science Reports**

**Decreased Atherosclerotic Lesion Formation in Human Serum Paraoxonase Transgenic Mice**

Aaron Tward, MA; Yu-Rong Xia, BS; Xu-Ping Wang, MD; Yi-Shou Shi, BS; Christina Park, BS; Lawrence W. Castellani, PhD; Aldons J. Lusis, PhD; Diana M. Shih, PhD

---

Received March 15, 2002; revision received May 4, 2002; accepted May 6, 2002.

From the Department of Medicine (A.T., Y.-R.X., X.-P.W., Y.-S.S., C.P., L.W.C., A.J.L., D.M.S.), the Department of Microbiology, Immunology, and Molecular Genetics (A.T., A.J.L.), and the Department of Human Genetics (A.J.L.), University of California, Los Angeles. A. Tward is now at the University of California, San Francisco.

Correspondence to Dr Diana M. Shih, Division of Cardiology, Department of Medicine, University of California, Los Angeles, 47-123 CHS, UCLA, Los Angeles, CA 90095-1679. E-mail dshih@mednet.ucla.edu

© 2002 American Heart Association, Inc.

Circulation is available at http://www.circulationaha.org

DOI: 10.1161/01.CIR.0000023623.87083.4F
Mice, Diets, and Atherosclerotic Lesion Analysis
Mice on the B6 background were maintained either on a 6% fat chow diet or, in the case of atherosclerotic lesion analysis, on a high fat diet (Teklad) containing 15.75% fat, 1.25% cholesterol, and 0.5% sodium chloride for 15 weeks. PON1 Tg mice on the B6 background were also backcrossed twice with the apoE KO mice on the B6 background (The Jackson Laboratory, Bar Harbor, Me) to obtain PON1 Tg/apoE KO mice and apoE KO littermates. These were maintained on a 6% fat chow diet. Lesions were analyzed as described.14

Southern Blot and PCR Analyses
A 32P-labeled human PON1 cDNA clone was used as a probe for Southern hybridization, and bands were visualized and quantified by using a PhosphorImager 445SI (Molecular Dynamics). For routine Southern blot analysis of PON1 Tg mice, PCR analysis was performed with the use of the primers 5′-GCTTGATTITTTCTCCAT-3′ and 5′-ATCTGTAATGTGCTAATCC-3′, yielding a 194-bp product.

Assays and LDL Oxidation
Mice were fasted for 16 hours before bleeding. Plasma lipids were determined by enzymatic colorimetric assays.14 Plasma PON1 and arylesterase activity assays and immunoblotting of human PON1 were performed as described.13 Plasma glucose levels were determined by using the glucose (Trinder) kit (Sigma-Aldrich). Mouse HDL was isolated in the absence of EDTA by ultracentrifugation as described.14 Human LDL was isolated by ultracentrifugation as described.14 For the LDL oxidation assay, human LDL (1 mg/mL in PBS) was incubated with 5 μmol/L CuSO4, with or without the presence of 0.25 mg/mL or 0.5 mg/mL mouse HDL, for 3 or 6 hours at 37°C. After the incubation, BHT was added to a final concentration of 20 μmol/L to stop the reaction. Lipid hydroperoxide contents of samples were then determined by use of the Fox assay.13

Statistical Analyses
The Student t test was used for analyzing all experimental data except for LDL oxidation, for which ANOVA and Fisher’s protected least significant difference (PLSD) test were used.

Results
Human PON1 Tg Mice
A 45-kb fragment containing the intact human PON1 gene with 10 kb of 5′-flanking and 10 kb of 3′-flanking sequences (Figure 1A) was used for the production of Tg mice. One Tg mouse line, designated PON1 Tg, carried ≈3 copies of the transgene, as determined by Southern blot analysis (data not shown). Tg mice were healthy and had normal weight, and the transgene was transmitted in a mendelian fashion. The PON1 transgene was maintained at a hemizygous state throughout the study so that insertional mutagenesis of the mouse genome was unlikely to contribute to the observed phenotypes, and studies with a separate line gave similar results (data not shown).

Northern blot analysis revealed that the transgene was expressed primarily in the liver (Figure 1B), exhibiting the same expression pattern as the endogenous mouse PON1 gene. Western blot analysis also confirmed the presence of human PON1 protein in the plasma of Tg mice (Figure 1C). By use of the arylesterase activity assay, plasma PON1 activities of the PON1 Tg mice on chow and high fat diets were 616 and 471 mOD270 · min⁻¹ · μL⁻¹, respectively, whereas plasma PON1 activities of the wild-type littermates on chow and high fat diets were 280 and 124 mOD270 · min⁻¹ · μL⁻¹, respectively (mOD270=absorbance [in milli-optical density] at wavelength 270 nm). Therefore, high fat diet feeding reduced plasma total PON1 (arylesterase) activities of the PON1 Tg and wild-type mice by 24% and 56%, respectively. Thus, when mice were maintained on a chow or a high fat diet, plasma PON1 (arylesterase) activities of the PON1 Tg mice were 2.2- and 3.8-fold higher, respectively, than those of the wild-type littermates on the same diet (Figure 2A and 2B). Hepatic mouse and human mRNA levels were also examined by Northern blot analysis in mice fed the 2 diets. Although expression of the mouse mRNA was decreased >50% in response to the high fat diet, expression of the human mRNA was not decreased by the same diet (Figure 2C).
Atherosclerotic Lesion Formation on B6 Background

Atherosclerotic lesion formation was examined in the PON1 Tg and wild-type littermates on the B6 background that were fed a high fat diet for 15 weeks. Compared with the female non-Tg littermates, the female Tg mice exhibited a 60% decrease in atherosclerotic lesion size ($P_{\text{H11005}}=0.13$, Figure 3A). The lack of a statistically significant result in the female group likely represents an effect of sample size. Compared with the male non-Tg littermates, the male Tg mice, on the other hand, exhibited a statistically significant 54% reduction in lesion size ($P_{\text{H11005}}=0.001$, Figure 3B). When data from the female and male mice were combined, the PON1 Tg mice, compared with the wild-type littermates, exhibited a statistically significant (57%) decrease in atherosclerotic lesion size ($P_{\text{H11005}}=0.01$, Figure 3C). Both the male and female PON1 Tg mice, compared with their wild-type littermates, exhibited similar plasma total cholesterol, VLDL/LDL cholesterol, HDL cholesterol, triglyceride, and glucose levels when they were maintained on either the chow or high fat diet (Table 1).

Atherosclerotic Lesion Formation on ApoE KO Background

The PON1 Tg mice were crossed onto the apoE KO genetic background for the study of advanced atherogenesis. When fed a chow diet, the PON1 Tg/apoE KO mice, compared with their apoE KO littermates, exhibited 2.5-fold higher plasma PON1 (arylesterase) activities (Table 2), whereas plasma total cholesterol, HDL cholesterol, VLDL/LDL cholesterol, triglyceride, and glucose levels were the same between the 2 groups (Table 2). Atherosclerotic lesion sizes of PON1 Tg/apoE KO mice at 6.5 months of age, on a chow diet, were 22% smaller than those of their apoE KO littermates ($P_{\text{H11005}}=0.01$) (Figure 4A). Aortic MCP-1 expression was also reduced by 44% in the PON1 Tg/apoE KO mice compared with their apoE KO littermates ($P_{\text{H11005}}=0.05$) (Figure 4B).
In Vitro Oxidation of LDL

Previously, we observed that PON1-deficient HDL lacked the ability to prevent LDL oxidation in a cell culture system of the arterial wall. In the present study, we examined the ability of HDLs isolated from the human PON1 Tg and wild-type mice on the B6 background to prevent LDL oxidation induced by copper in vitro. For HDL isolated from mice on the B6 background, we found that the PON1 Tg HDL, compared with the wild-type HDL, exhibited 3.2-fold higher arylesterase activity (152,000 versus 48,000 mOD min⁻¹ mg⁻¹ protein⁻¹). At a dose of 0.25 mg HDL/mL, compared with the LDL plus wild-type HDL group, the LDL plus PON1 Tg HDL group exhibited statistically significant 33% (P=0.0001, Fisher’s PLSD) and 14% (P=0.001, Fisher’s PLSD) decrease in lipid hydroperoxide level (Figure 6A). After 6 hours of incubation and at the same dose of 0.25 mg HDL/mL, compared with the LDL group, neither the LDL plus PON1 Tg/apoE KO HDL group nor the LDL plus apoE KO HDL group exhibited any significant reduction in lipid hydroperoxide level (Figure 6A). At a higher dose of 0.5 mg HDL/mL, the LDL plus PON1 Tg/apoE KO HDL group, compared with the LDL plus apoE KO HDL group, exhibited statistically significant 21% (P<0.0001, Fisher’s PLSD) and 14% (P<0.01) reduction in lipid hydroperoxide levels after 3 and 6 hours of incubation, respectively (Figure 6B). Thus, on the apoE KO mouse background, PON1 Tg HDL was also more effective in preventing LDL oxidation than was wild-type HDL.

Discussion

Both biochemical and epidemiological studies suggest that LDL oxidation plays a major role in the development of atherosclerosis. However, human clinical trials and animal studies that used antioxidant supplementation to treat cardiovascular disease have been inconclusive. Alternative targets for the prevention and/or treatment of atherosclerosis

| Table 1. Plasma Lipid and Glucose Levels of PON1 Tg and Wild-Type Mice Fed Chow or High-Fat Diet |

<table>
<thead>
<tr>
<th>Mice</th>
<th>Sample Size, n</th>
<th>Total Cholesterol, mg/dL</th>
<th>VLDL/LDL Cholesterol, mg/dL</th>
<th>HDL Cholesterol, mg/dL</th>
<th>Triglycerides, mg/dL</th>
<th>Glucose, mg/dL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Female, chow diet</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wild type</td>
<td>15</td>
<td>97±4</td>
<td>19±3</td>
<td>77±2</td>
<td>20±6</td>
<td>100±7</td>
</tr>
<tr>
<td>PON1 Tg</td>
<td>14</td>
<td>90±4</td>
<td>18±3</td>
<td>72±2</td>
<td>22±8</td>
<td>108±3</td>
</tr>
<tr>
<td>Male, chow diet</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wild type</td>
<td>19</td>
<td>105±2</td>
<td>19±2</td>
<td>86±2</td>
<td>42±6</td>
<td>115±4</td>
</tr>
<tr>
<td>PON1 Tg</td>
<td>23</td>
<td>107±2</td>
<td>18±1</td>
<td>88±1</td>
<td>35±3</td>
<td>114±4</td>
</tr>
<tr>
<td>Female, high-fat diet</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wild type</td>
<td>13</td>
<td>277±19</td>
<td>219±20</td>
<td>58±6</td>
<td>5±1</td>
<td>106±5</td>
</tr>
<tr>
<td>PON1 Tg</td>
<td>10</td>
<td>308±31</td>
<td>253±34</td>
<td>56±6</td>
<td>5±1</td>
<td>106±7</td>
</tr>
<tr>
<td>Male, high-fat diet</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wild type</td>
<td>19</td>
<td>320±17</td>
<td>249±19</td>
<td>71±3</td>
<td>6±1</td>
<td>124±5</td>
</tr>
<tr>
<td>PON1 Tg</td>
<td>21</td>
<td>318±16</td>
<td>243±18</td>
<td>75±3</td>
<td>8±1</td>
<td>126±5</td>
</tr>
</tbody>
</table>

Data are mean±SEM.

*P<0.0001 vs corresponding value for apoE KO mice.

| Table 2. Plasma Arylesterase Activity, Glucose, and Lipid Levels of PON1 Tg/ApoE KO Mice and ApoE KO Mice Fed a Chow Diet |

<table>
<thead>
<tr>
<th>Mice</th>
<th>Sample Size, n</th>
<th>Arylesterase Activity, mOD₂₇₀ min⁻¹ µL</th>
<th>Total Cholesterol, mg/dL</th>
<th>VLDL/LDL Cholesterol, mg/dL</th>
<th>HDL Cholesterol, mg/dL</th>
<th>Triglycerides, mg/dL</th>
<th>Glucose, mg/dL</th>
</tr>
</thead>
<tbody>
<tr>
<td>ApoE KO</td>
<td>18</td>
<td>247±13</td>
<td>486±47</td>
<td>461±48</td>
<td>25±3</td>
<td>57±4</td>
<td>159±16</td>
</tr>
<tr>
<td>PON1 Tg/ApoE KO</td>
<td>18</td>
<td>610±42*</td>
<td>472±24</td>
<td>445±23</td>
<td>27±2</td>
<td>56±5</td>
<td>144±11</td>
</tr>
</tbody>
</table>

Data are mean±SEM.

*P<0.0001 vs corresponding value for apoE KO mice.
are antioxidative enzymes on HDL, such as PON1. We have previously shown that mice lacking PON1 had increased atherogenesis, which is consistent with predictions from in vitro studies. To determine whether high levels of PON1 can protect against LDL oxidation and decrease atherosclerosis in vivo, we have generated and characterized human PON1 Tg mice. Our results, indicating protection against both early and late stages of atherogenesis, are significant in that they demonstrate a dose-dependent effect of PON1. Hence, normal levels of PON1 are limiting with respect to their antiatherosclerotic activity. The present study is also the first to demonstrate that the human PON1 protein exhibits antioxidative and antiatherogenic functions in vivo.

We observed that the human PON1 transgene was expressed in a liver-specific manner (Figure 1), similar to the expression pattern of the endogenous mouse Pon1 gene, suggesting that the transgene constructs contain the necessary cis-regulating elements for liver-specific expression. Interestingly, expression of the human PON1 mRNA, unlike the endogenous mouse Pon1 mRNA, was not decreased by the high fat diet (Figure 2C). It is unclear whether this represents a true difference between human and mouse regulation of PON1 or if it is simply due to the construct that we used. On the basis of Northern blot analysis (Figure 2C), we conclude that the high fat diet–induced reduction of plasma total PON1 activities in the PON1 Tg and wild-type mice (Figure 2B versus 2A) is probably caused by the decreased mouse Pon1 mRNA levels.

We observed that HDL isolated from the PON1 Tg mice, compared with the wild-type HDL, exhibited ~3-fold higher PON1 activity and was more effective at protecting against LDL oxidation (Figures 5 and 6). Several antioxidative functions of PON1 have been elucidated and include a phospholipase A2–like activity that hydrolyzes biologically active oxidized phospholipids (such as phosphatidylcholine...
isoprostane and core aldehydes), and peroxidase-like activities that destroy lipid hydroperoxides, and H₂O₂. Thus, PON1 appears to exert its antiatherogenic effects mainly through its antioxidative functions. In addition to preventing LDL oxidation, PON1 may preserve other functions of HDL, such as reverse cholesterol transport, by reducing oxidative damage to HDL. We also observed a 44% reduction of aortic MCP-1 expression in the PON1 Tg/apoE KO mice compared with their apoE KO littermates (Figure 4B). MCP-1, a chemokine induced by minimally oxidized LDL, plays a key role in monocyte recruitment and fatty streak formation. Deficiency in MCP-1 or its receptor, chemokine receptor-2, resulted in reduced atherosclerotic lesion formation in genetically altered mice. Our results are consistent with an antiatherosclerotic role for PON1 upstream from MCP-1 induction, most likely by increasing the antioxidative activity of HDL and subsequently reducing the levels of oxidized LDL in the arterial wall.

Human epidemiological studies have revealed an association between low PON1 levels and an increased risk for coronary artery disease. In the present study, we observed that a moderate increase in PON1 levels protects against atherosclerotic lesion formation in the PON1 Tg mice, compared with the wild-type mice, at both the early fatty streak stage (B6 model) (Figure 3) and the intermediate- to advanced-lesion stage (apoE KO model) (Figure 4A). Thus, PON1 may be a potential therapeutic agent for the prevention and/or treatment of atherosclerosis. Consistent with its ability to reduce cardiovascular mortality, a recent study has demonstrated that moderate alcohol consumption increases plasma HDL, apoA-I, and PON1 levels in volunteers. Also, cigarette smoking is associated with reduced serum PON1 activity. Interestingly, flavonoids, such as glabridin (found in licorice) and quercetin (found in red wine), have been shown to scavenge reactive oxygen species and preserve the anti-oxidative functions of PON1 in vitro. Supplementation of flavonoids protects LDL against oxidation and attenuates atherosclerosis in animal models. Therefore, it will be interesting to examine whether a combined supplementation of flavonoids (or other antioxidants) and PON1 will exert a synergistic effect on protection against LDL oxidation and atherosclerosis in animal models. Further analysis of the regulation of PON1 levels in humans may yield promising avenues for atherosclerosis therapy.

Acknowledgments

This work was supported by the US Public Health Service (grant HL-30568 to Dr Lusis) and by the American Heart Association (Western States Affiliate Grant 0060040Y to Dr Shih). We thank Dr Clement Furlong for providing PON1 cDNA and antibody and Thomas J. Fielder for DNA microinjection. We thank Janet Daniger, Sharda Chaugundla, Jack Wong, and John Miller for excellent technical assistance.

References


Decreased Atherosclerotic Lesion Formation in Human Serum Paraoxonase Transgenic Mice
Aaron Tward, Yu-Rong Xia, Xu-Ping Wang, Yi-Shou Shi, Christina Park, Lawrence W. Castellani, Aldons J. Lusis and Diana M. Shih

Circulation. 2002;106:484-490; originally published online July 1, 2002; doi: 10.1161/01.CIR.0000023623.87083.4F
Circulation is published by the American Heart Association, 7272 Greenville Avenue, Dallas, TX 75231
Copyright © 2002 American Heart Association, Inc. All rights reserved.
Print ISSN: 0009-7322. Online ISSN: 1524-4539

The online version of this article, along with updated information and services, is located on the
World Wide Web at:
http://circ.ahajournals.org/content/106/4/484

Permissions: Requests for permissions to reproduce figures, tables, or portions of articles originally published in Circulation can be obtained via RightsLink, a service of the Copyright Clearance Center, not the Editorial Office. Once the online version of the published article for which permission is being requested is located, click Request Permissions in the middle column of the Web page under Services. Further information about this process is available in the Permissions and Rights Question and Answer document.

Reprints: Information about reprints can be found online at:
http://www.lww.com/reprints

Subscriptions: Information about subscribing to Circulation is online at:
http://circ.ahajournals.org/subscriptions/